

Investigation of the usability of RIKEN ^{44m}Sc for radiolabeling on chelate-compounds

S. Oshikiri,^{*1,*2} S. Kubota,^{*1,*2} H. Kato,^{*1,*2} T. Yokokita,^{*1} K. Suzuki,^{*1,*2} A. Hino,^{*2} and H. Haba^{*1}

Scandium-44m (^{44m}Sc) with a half-life of 58.61 h decays to scandium-44 (^{44}Sc) with a half-life of 3.97 h by emitting gamma rays. ^{44}Sc has been reported to be a promising radioisotope (RI) in positron emission tomography imaging, while ^{47}Sc , which emits a beta particle, is proposed as a promising therapeutic nuclide.¹⁾ Furthermore, ^{44m}Sc appears to be useful as an imaging RI²⁾ and a surrogate nuclide for other Sc isotopes in basic science research because of a relatively long half-life. A study on the optimization of ^{44m}Sc radiolabeling of macrocyclics-functionalized biomolecules has been published.³⁾

We have been developing a production method of ^{44m}Sc via the nuclear reaction $^{44}\text{Ca}(d, 2n)^{44m}\text{Sc}$ at the RIKEN AVF cyclotron and distributing purified ^{44m}Sc (RIKEN ^{44m}Sc) to users.⁴⁾ To confirm the quality of the RIKEN ^{44m}Sc for research on radiolabeling, we performed ^{44m}Sc radiolabeling on chelate-compounds in this study. To investigate the structural effect of chemical-compounds on ^{44m}Sc radiolabeling, four commercially available compounds were selected: DOTA-Substance P (DOTA-SP), DOTA-RGD₂, NOTA-RGD₂ and NODAGA-RGD₂. The operations for ^{44m}Sc radiolabeling are as described below.

- Step 1: RIKEN ^{44m}Sc (9.1 MBq) was dissolved in 0.05 M hydrochloric acid to prepare a ^{44m}Sc solution (79 MBq/mL). The radioactivity of ^{44m}Sc was determined using a germanium semiconductor detector.
- Step 2: Each chelate-compound was dissolved in 0.75 M sodium acetate buffer at pH3.0, 4.0, 5.0, and 6.0 to prepare 1.4×10^{-4} M sample solutions.
- Step 3: 1.5 μL of the ^{44m}Sc solution was added to 3 μL of each sample solution: the specific radioactivity of each sample solution was 0.29 MBq/nmol.
- Step 4: The mixtures in Step 3 were heated at 97°C for 10 min and kept at 20°C for 5 min.
- Step 5: Radiolabeling yields of ^{44m}Sc -labeled compounds were determined using thin-layer chromatography (TLC) with a C18 reverse phase TLC plate (NAGEL RP-18W/UV254), which was eluted with a mixture of acetonitrile, 0.5 M ammonium acetate, methanol, and tetrahydrofuran in a volume ratio of 4 : 3 : 2 : 1 using an image analyzer.

Consequently, the radiolabeling yields of each compound were over 90% at pH 5.0–6.0, although they were different from each other at lower pH values. In case of compounds with the same affinity moiety, DOTA-

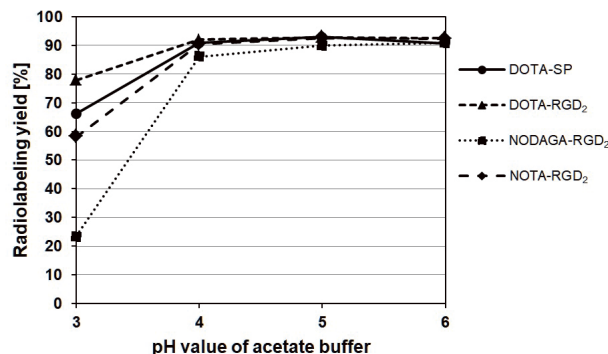


Fig. 1. Relation between pH value of acetate buffer and radiolabeling yield (%) of ^{44m}Sc -labeled compounds at 0.29 MBq/nmol ($n = 1$).

RGD₂ was radiolabeled over 70% at pH 3.0 which was higher than that of NODAGA-RGD₂ and NOTA-RGD₂.

It has been reported that the suitable pH range for ^{44m}Sc labeling with DOTA-based ligands is from 4 to 5.5.³⁾ The result obtained in our study was consistent with the result of the previous study. Moreover, regarding the difference in reactivity among chelators, one report shows that DOTA is a better chelator than NODAGA because NODAGA is more susceptible to contamination metals than DOTA.⁵⁾ Another report shows that the thermodynamic stability of Sc-DOTA is high compared to that of Sc-NOTA.⁶⁾ These reports are consistent with our radiolabeled result at pH 3.0. Hence, these results support the possibility of further radiolabeling studies using RIKEN ^{44m}Sc .

We investigated the possibility of RIKEN ^{44m}Sc for radiolabeling studies and compared our results with those of previous reports of Sc radioisotopes. In addition, the pH responsiveness of the ^{44m}Sc radiolabeling yield to compounds having different chelating sites was confirmed. In future, we plan to optimize the radiolabeling condition of Sc isotopes, such as ^{44}Sc , which is eluted from the $^{44}\text{Ti}/^{44}\text{Sc}$ generator, and conduct a feasibility study for imaging tracer with ^{44m}Sc .

References

- 1) C. Müller *et al.*, J. Nucl. Med. **55**, 1658 (2014).
- 2) T. Fukuchi *et al.*, RIKEN Accel. Prog. Rep. **53**, 21 (2020).
- 3) S. Huclier-Markai *et al.*, Nucl. Med. Biol. **41**, e36 (2014).
- 4) H. Haba, Drug Deliv. Syst. **35**, 114 (2020).
- 5) K. A. Domnanich *et al.*, EJNMMI Radiopharm. Chem. **1**, 8 (2016).
- 6) S. Huclier-Markai *et al.*, Radiochim. Acta **99**, 653 (2011).

*1 RIKEN Nishina Center

*2 RI Research Department, FUJIFILM Toyama Chemical Co., Ltd.